

LAVORO ORIGINALE

Blocking TNF in vitro with infliximab determines the inhibition of expansion and interferon gamma production of V γ 9/V δ 2 T lymphocytes from patients with active rheumatoid arthritis.

A role in the susceptibility to tuberculosis?*

*Il blocco del TNF in vitro determina inibizione della espansione e della produzione di interferone gamma da parte di linfociti V γ 9/V δ 2 di pazienti con artrite reumatoide. Un ruolo nella suscettibilità alla tubercolosi?**

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RIASSUNTO

Tra gli effetti collaterali della neutralizzazione del TNF, per la maggior parte di tipo infettivo, il più importante appare la riattivazione di un processo tubercolare. Il subset di linfociti $\gamma\delta$ sembra giocare un ruolo importante nella protezione contro il bacillo tubercolare.

Oggetto dello studio è stato quello di valutare la capacità di espansione di tale subset linfocitario in pazienti con artrite reumatoide (AR) e di studiare l'effetto in vitro di infliximab.

Sono stati inclusi nello studio 42 pazienti affetti da AR (28 PPD positivi, 14 PPD negativi) e 155 controlli (110 PPD positivi e 45 PPD negativi) ed è stata valutata la capacità di espansione, l'espressione del recettore per il TNF e il contenuto di Interferone dei linfociti $\gamma\delta$.

Il fattore di espansione dei linfociti $\gamma\delta$ risultò più elevato nei soggetti PPD positivi, con i livelli più alti nei pazienti affetti da AR. L'aggiunta di infliximab nelle culture di linfociti $\gamma\delta$ dei pazienti PPD positivi determinò una significativa inibizione dell'espansione, della espressione del recettore del TNF e del contenuto di Interferone γ .

Conclusioni: I soggetti con artrite reumatoide PPD positiva hanno una elevata capacità proliferativa del subset $\gamma\delta$ in risposta ai fosfoantigeni che è inibita in vitro dalla esposizione a infliximab. È ipotizzabile che il blocco del TNF può svolgere un ruolo nella emergenza dell'infezione tubercolare attraverso l'inibizione di tale subset.

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INTRODUCTION

Biological therapeutic agents neutralising tumour necrosis factor (TNF) are highly active in treating chronic inflammatory diseases, such as

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Crohn's disease, rheumatoid arthritis, ankylosing spondylitis, uveitis, and psoriasis (1-3). From the beginning, side effects of TNF neutralisation - mostly infectious complications - were recognized, the most important being, however, pulmonary tuberculosis infections (4).

TNF is a cytokine produced primarily by macrophages in response to stimuli activating toll-like receptors, but can also be expressed by activated T cells, B cells, and NK cells (5, 6). An effective host response against mycobacterium tuberculosis (Mtb) involves the differentiation of specific T cells to secrete an appropriate Th1 cytokine pro-

file and the development of granulomas in which activated epithelioid macrophages restrict mycobacterial growth and TNF is necessary for optimal coordination of both aspects of Mtb immunity (7).

The peripheral blood of patients with rheumatoid arthritis (RA) contains oligoclonal gamma/delta T cell populations which may contribute to the pathogenesis of the disease (8, 9).

Gamma/delta T cells and dendritic cells are quickly recruited to the lungs shortly after intranasal vaccination with BCG and can contribute to protective immune response against *Mycobacterium tuberculosis* (10). It has been also previously reported that V γ 9/V δ 2 T cells from tuberculin purified protein derivative (PPD)-positive children, either healthy or affected by different clinical forms of tuberculosis, strongly proliferate to different phosphoantigens *in vitro*, whereas V γ 9/V δ 2 T cells from PPD-negative healthy subjects proliferate very poorly (11, 12). Disease-associated changes in V γ 9/V δ 2 T cell effector functions in patients with tuberculosis are consistent with the possibility that these T cells may play a protective role in immune response against *M. tuberculosis* infection.

Aim of the present study was to assess the expansion capacity of V γ 9/V δ 2 T cells from (PPD positive and PPD negative) patients with active rheumatoid arthritis, and to examine the *in vitro* effect of infliximab on this lymphocyte subset.

MATERIALS AND METHODS

Patients

Forty-two patients with rheumatoid arthritis classified according to the 1987 revised ACR criteria (13) were studied. Twenty-eight were PPD-negative (8 males and 20 females, mean age 57 \pm 12 years), 14 were PPD-positive (4 males, 10 females, mean age 64 \pm 7 years). At time of sampling, the disease was active in all patients. Treatments included methotrexate (10-20 mg per week) with or without NSAID and/or steroids (prednisone <10 mg/day). Forty-five PPD-negative (mean age 34 \pm 8 years) and 110 PPD-positive (mean age 39 \pm 12 years) healthy volunteers were enrolled as controls. Human studies committee approval and individual informed consent from each subject were obtained.

Monoclonal antibodies and flow cytometry

mAbs specific for human surface antigens anti-CD3 phycoerythrin-labelled (PE) and anti-T-cell

receptor (TCR) V δ 2 fluorescein isothiocyanate (FITC; PharMigen, San Diego, CA, USA) were used as follows. Peripheral blood mononuclear cells (PBMCs; 10⁶ in 100 μ l phosphate buffered saline with 1% heat-inactivated foetal calf serum and 0.02% Na-azide) were incubated at 4°C for 30 min with anti-CD3-PE conjugated mAb and anti-TCR V δ 2 FITC conjugated mAb simultaneously. After washing, the cells were suspended in PBS with 1% foetal calf serum and analyzed on a FAC-Scan flow cytometer (Becton Dickinson, Mountain View, CA, USA) by using forward scatter/side scatter gating to select the lymphocyte population for analysis.

Cell separation and expansion in vitro of V γ 9/V δ 2 T lymphocytes

PBMCs were obtained from each individual by separating heparinized venous blood on Ficoll (Euroclone, Wetherby, Yorkshire, UK). The cells were washed in RPMI-1640 medium (Euroclone), and cultured in 24-well plates (Costar, Cambridge, MA, USA) at a concentration of 5 \times 10⁵ cells/ml in RPMI 1640 supplemented with 10% foetal calf serum (Euroclone), hepes 20 mmol/l (Euroclone), 2 mmol/l L-glutamine (Euroclone) and penicillin/streptomycin 100 U/ml (Sigma, St Louis, USA), at 37°C and at 0.5% CO₂.

For the expansion of V γ 9/V δ 2 T cells, PBMCs were cultured for 10 days in medium alone or in the presence of the follow phosphoantigens: xylose 1-P (Sigma; 0.5 mmol/l final concentration); ribose 1-P (Sigma; 0.5 mmol/l final concentration); dimethylallyl pyrophosphate (DMAPP; Sigma; 0.5 mmol/l final concentration).

After 72 hours, cultures were supplemented with a 0.5 ml medium containing 20 U/ml recombinant human interleukin (IL-2; Genzyme, Cambridge, MA, USA). Every 72 hours, 0.5 medium was replaced with a 0.5 ml fresh medium containing 20 U/ml IL-2.

After 10 days, cells were washed three times in medium, and expansion of V γ 9/V δ 2 T cells was assessed using FACScan, as described above.

The absolute number of V γ 9/V δ 2 T cells in each culture was calculated according to the following formula: % V γ 9/V δ 2 positive cells before culture \times total cell count/100. The V γ 9/V δ 2 expansion factor (EF) was then calculated by dividing the absolute number of V γ 9/V δ 2 T cells in specifically stimulated cultures by the absolute number of V γ 9/V δ 2 T cells cultured in the absence of any antigen.

In vitro effect of infliximab on V γ 9/V δ 2 T lymphocyte cultures

In order to examine the effects on V γ 9/V δ 2 T lymphocyte expansion, TNF-RII expression and IFN- γ content, infliximab (Remicade; Centocor Inc., Malvern, PA; Shering Plough SpA, Italy) was added in the medium at a final concentration of 10, 50 (for 3 days) and 100 μ g/ml (for 10 days). The surface expression of TNF-RII expression was studied using anti-TNF receptor II PE (R&D system, Minneapolis, MN; USA) and anti V δ 2 simultaneously. The number of TNF-RII molecules (MESF; molecular equivalents of soluble fluorochrome) was calculated by FACS analysis of cells stained with saturating amounts of PE labelled anti-TNF-RII of known PE/protein ratio and comparing the staining with a standard curve of microbeads labelled with defined numbers of PE molecules (Quantum Fluorescence Kit, Sigma). The analysis was done using Quickcal Program for MESF Units for Windows. For the evaluation of intracytoplasmic content of IFN γ , 3×10^5 cells were stained with anti-V δ 2 TCR FITC and, after washing, fixed with 4% paraformaldehyde (Sigma) for 30 minutes at 4°C. After two washes with permeabilization buffer (saponine containing) the cells were incubated at 4°C for 45 minutes with anti-IFN- γ PE (Euroclone). After washing, the cells were suspended in PBS with 1% foetal calf serum and data were acquired on a FACScan instrument and analyzed using WinMDI version 2.8 software.

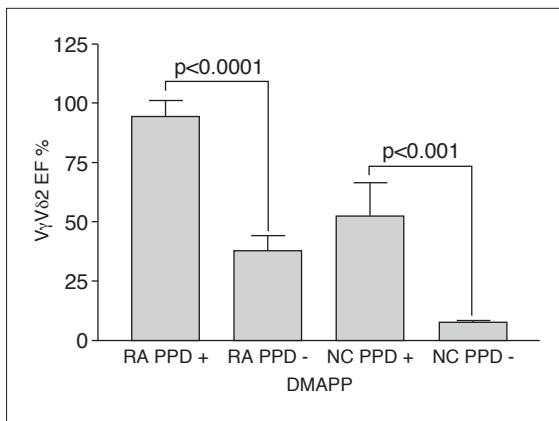


Figure 1 - Expansion of V γ 9/V δ 2 T lymphocytes from patients with PPD positive and negative rheumatoid arthritis and PPD positive and negative controls in response to dimethylallylpyrophosphate (DMAPP). The V γ 9/V δ 2 T cell expansion factor (EF) was calculated by dividing the absolute number of V γ 9/V δ 2 T cell in specifically DMAPP stimulated cultures by the absolute number of V γ 9/V δ 2 T cells cultured in the absence of any antigen. Results are expressed as mean \pm SD.

STATISTICAL ANALYSIS

Kruskal-Wallis test followed by a Dunnet's post hoc analysis was used to compare continuous variables in the different groups while Wilcoxon rank test was used to evaluate effect of infliximab on cell expansion, TNFRII expression and IFN γ content. A p value <0.05 was considered significant.

RESULTS

Expression of V γ 9/V δ 2 T-cell receptor on lymphocytes in peripheral blood

The percentage of gamma/delta T cells with phenotype V γ 9/V δ 2 was similar in both patients and normal individuals ($4.38 \pm 1.56\%$ and $3.05 \pm 1.34\%$, respectively).

The number of circulating V γ 9/V δ 2 T cells also was not substantially modified by different therapies or by the presence of PPD positivity.

Expansion in vitro of V γ 9/V δ 2 T cells

The expansion of V γ 9/V δ 2 T lymphocytes was evaluated *in vitro* by incubating the cells with five DMAPP for 10 days or in medium (containing IL-2) alone.

At this time the percentage of expansion was assessed by fluorescence activated cell sorting (FACS) analysis using the anti-TCR V δ 2 mAb. The results were expressed as EF (see materials and

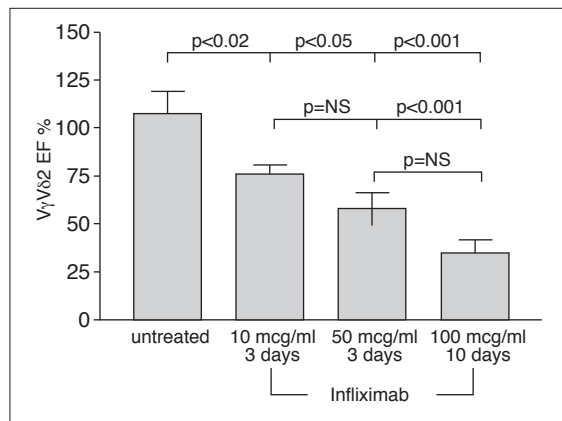


Figure 2 - *In vitro* effect of infliximab incubation on V γ 9/V δ 2 T lymphocytes expansion (EF) from patients with PPD positive rheumatoid arthritis. Results are expressed as mean \pm SD.

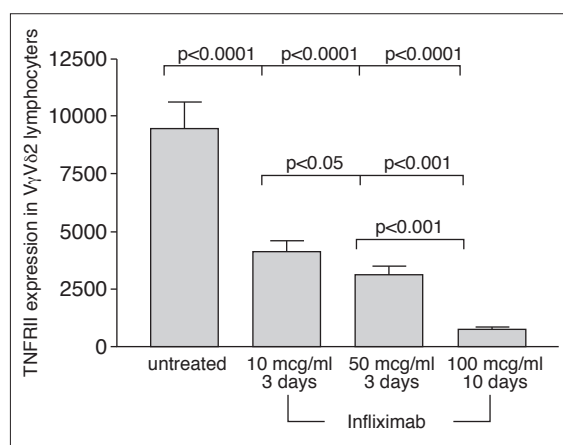


Figure 3 - Effect of infliximab on TNFRII expression of Vγ9/Vδ2 T lymphocytes from patients with PPD positive rheumatoid arthritis. Levels were expressed as MESF (molecular equivalents of soluble fluorochrome). Results are expressed as mean ± SD.

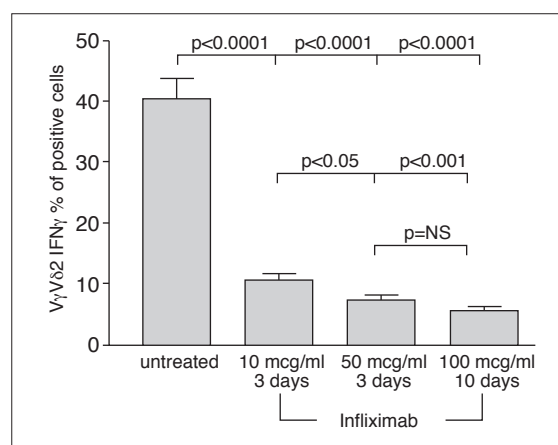


Figure 4 - Effect of infliximab on IFN-gamma content of Vγ9/Vδ2 T lymphocytes from patients with PPD positive rheumatoid arthritis. Results are expressed as mean ± SD of percentage of positive cells.

methods). Significant differences were found in the response to DMAPP in the tested groups (Fig. 1). Specifically, the EF of Vγ9/Vδ2 T cells of RA patients was higher than that of healthy control individuals, and differences were observed between PPD-positive and negative patients and controls (RA PPD-positive 96 ± 38 vs RA PPD-negative 38 ± 26 , $p < 0.01$; NC PPD-positive 51 ± 98 , vs NC PPD-negative 8 ± 5.8 , $p < 0.01$; RA PPD positive vs NC PPD positive, NS).

In vitro studies with infliximab

The addition of infliximab to the cultures from 7 PPD positive-patients determined a significant inhibition of cell expansion (108 ± 27 in untreated cultures, 76 ± 12 in cultures treated for 3 days with $10 \mu\text{g/ml}$ infliximab, 58 ± 20 in cultures treated for 3 days with $50 \mu\text{g/ml}$ and 35 ± 18 in those treated for 10 days with $100 \mu\text{g/ml}$ infliximab) (Fig. 2) and a significant decrease of TNF-RII expression (Fig. 3).

In particular, the mean TNF-RII MESF was 10.422 ± 1.694 in untreated culture and 4.561 ± 615 (infliximab $10 \mu\text{g/ml}$ for 3 days), 3.422 ± 584 (infliximab $50 \mu\text{g/ml}$ for 3 days) and 820 ± 175 (infliximab $100 \mu\text{g/ml}$ for 10 days) respectively after exposure to infliximab.

IFNγ (Fig. 4) in untreated cultures was $40.4 \pm 8.2\%$. In the presence of infliximab we found values of $10.5 \pm 3\%$ (infliximab $10 \mu\text{g/ml}$ for 3 days), $7.2 \pm 2.2\%$ (infliximab $50 \mu\text{g/ml}$ for 3 days) and $5.5 \pm 1.7\%$ (infliximab $100 \mu\text{g/ml}$ for 10 days) respectively.

DISCUSSION

The use of TNF antagonists is accompanied by an increased risk of active tuberculosis. Although the risk seems to pertain to all 3 available agents, it has been suggested that infliximab and adalimumab carry a higher risk than etanercept (14).

The role of TNF in the human immune response to tuberculosis remains unclear. This cytokine may be responsible for some of the clinical manifestations of the disease, including weight loss, night sweats, and tissue destruction (15). Membrane-bound TNFα seems to provide major protection against Mycobacterium infection in mice models. Yet in animal models, TNFα plays a central part in the host response against tuberculosis, including granuloma formation and containment of disease. Antibodies against TNFα cause a reactivation of tuberculosis in a mouse model of latent infection. Given the key role of TNFα in the innate immune response to tuberculosis, patients receiving treatment with infliximab are probably also susceptible to disease after primary infection and exogenous reinfection with Mtb (16).

In the present study we analyzed the *in vitro* expansion capacity of Vγ9/Vδ2 T cells from patients with rheumatoid arthritis.

Vγ9/Vδ2 T cells represent the majority of gamma/delta T lymphocytes in the peripheral blood (17) and phosphoantigens are known to activate specifically Vγ9/Vδ2 T cells in a major histocompatibility complex unrestricted, but TCR dependent, manner (18).

An increase in γ/δ T cell expansion was observed in RA patients when compared to healthy subjects, our results being not sufficient to hypothesize a role of this subset in the pathogenesis of the disease. Different results, indeed, have previously been reported, but this discrepancy is probably due to inclusion of different populations of patients and/or stages of disease progression (19, 20).

A high proportion of PPD positive RA patients had an increase in V γ 9/V δ 2 T cells expansion suggesting a role in the protection against M. Tuberculosis in these patients. V γ 9/V δ 2 T cells from patients, but not from control individuals, responded to DMAPP *in vitro* with expansion. This phenomenon might be explained by the fact that V γ 9/V δ 2 T cells from PPD positive patients are pre-activated *in vivo*. Alternatively, V γ 9/V δ 2 T cells in PPD positive patients might be less susceptible to apoptosis and account for the increased expansion.

In vitro exposure of gamma/delta T cells to infliximab determined a significant depression of γ/δ T cell expansion and IFN production. The effect of infliximab might explain the increase susceptibility of latent tuberculosis in RA patients to overt infection when exposed to TNF blocking agents.

Studies in humans and animal models have suggested in fact that γ/δ T cells may play an important role in the immune response to Mtb (10-13). In mice, there is a large increase in the number of γ/δ T cells that accumulate in the lungs after intranasal challenge with PPD and, in some experimental con-

ditions, mice lacking gamma/delta T cells suffer a more severe form of tuberculosis and fail to control the infection. In normal healthy individuals, γ/δ T cells contain the highest frequency of Mtb-reactive T cells in the peripheral blood and the predominant subset of Mtb-reactive γ/δ T cells express a TCR encoded by V γ 9 and V δ 2 gene segments (21). V γ 9/V δ 2 T lymphocytes reduce the viability of intracellular Mtb. Participation in early immunity against Mtb is also a feature of a subset of mouse T cells with the antigen receptor variable region encoded by the Vc1 gene. Vc1 cells are rapidly recruited to the lungs of mice infected intranasally with BCG, are able to produce TNF α and IFN γ following *in vitro* stimulation, and are cytotoxic against BCG-infected macrophages (22). We have also found that V γ 9/V δ 2 T cells from children with tuberculosis have an increased proliferative activity, but decreased IFN-gamma production and granulysin expression (23). After successful chemotherapy, the V γ 9/V δ 2 T cell proliferative response strongly decreased, whereas IFN γ and granulysin production consistently increased (23).

In conclusions in this study we have documented that γ/δ T lymphocytes from patients with PPD positive rheumatoid arthritis have a high capacity to respond *in vitro* to phosphoantigens with expansion and IFN γ production that is inhibited by the exposure to infliximab. These results might be of relevance in view of the effect of TNF blocking on the pulmonary tuberculosis infection.

SUMMARY

Background: Side effects of TNF neutralisation - mostly infectious complications - were recognized, the most important being pulmonary tuberculosis infections. γ/δ T cells contribute to protective immune response against mycobacterium tuberculosis.

Objectives: The aim of the present study was to assess the expansion capacity of V γ 9/V δ 2 T cells from (tuberculin purified protein derivative (PPD) positive and PPD negative) patients with active rheumatoid arthritis (RA), and to examine the *in vitro* effect of infliximab on this lymphocyte subset.

Methods: 28 PPD negative RA patients were studied and compared with 14 PPD positive RA patients, 45 PPD-negative and 110 PPD-positive healthy volunteers. Cell separation, expansion *in vitro* of V γ 9/V δ 2 T lymphocytes (EF) and the expression of tumor necrosis factor receptor II and IFN-gamma content by V γ 9/V δ 2 T lymphocytes were studied before and after infliximab *in vitro* addition.

Results: The EF from PPD positive subjects was higher than that from PPD negatives. Patients with RA have the highest levels. The addition of infliximab to the cultures from PPD-positive patients determined a significant inhibition of cell expansion and TNF RII expression and a significant decrease of IFN gamma content.

Conclusion: In this study we have documented that γ/δ T lymphocytes from patients with PPD positive rheumatoid arthritis have a high capacity to respond *in vitro* to phosphoantigens with expansion TNF-RII expression and IFN-gamma production that is inhibited by the exposure to infliximab. These results might be of relevance in view of the effect of TNF blocking on the pulmonary tuberculosis infection.

Parole chiave - Artrite reumatoide, infliximab, tubercolosi, linfociti γ/δ .

Key words - Rheumatoid arthritis, tuberculosis infection, infliximab, V γ 9/V δ 2 T lymphocytes.

REFERENCES

1. Sandborn WJ, Hanauer SB. Infliximab in the treatment of Crohn's disease: a user's guide for clinicians. *Am J Gastroenterol* 2002; 97: 2962-72.
2. Chaudhari U, Romano P, Mulcahy LD, Dooley LT, Baker DG, Gottlieb AB. Efficacy and safety of infliximab monotherapy for plaque-type psoriasis: a randomised trial. *Lancet* 2001; 357: 1842-7.
3. Criscione LG, St Clair EW. Tumor necrosis factor-alpha antagonists for the treatment of rheumatic diseases. *Curr Opin Rheumatol* 2002; 14: 204-11.
4. Keane J, Gershon S, Wise RP, Mirabile-Levens E, Kasznica J, Schwietzman WD, et al. Tuberculosis associated with infliximab, a tumor necrosis factor alpha-neutralizing agent. *N Engl J Med* 2001; 345: 1098-104.
5. Papadakis KA, Targan SR. Tumor necrosis factor: biology and therapeutic inhibitors. *Gastroenterology* 2000; 119: 1148-57.
6. Sedgwick JD, Riminton DS, Cyster JG, Korner H. Tumor necrosis factor: a master-regulator of leukocyte movement. *Immunol Today* 2000; 21: 110-13.
7. Roach DR, Bean AG, Demangel C, France MP, Briscoe H, Britton WJ. TNF regulates chemokine induction essential for cell recruitment, granuloma formation, and clearance of mycobacterial infection. *J Immunol* 2002; 168: 4620-7.
8. Olive C, Gatenby PA, Serjeantson SW. Persistence of gamma/delta oligoclonality in the peripheral blood of rheumatoid arthritis patients. *Immunol Cell Biol* 1994; 72: 7-11.
9. Bank I, Cohen L, Moullem M, Farfel Z, Grossman E, Ben-Nun A. $\gamma\delta$ T cell subsets in patients with arthritis and chronic neutropenia. *Ann Rheum Dis* 2002; 61: 438-43.
10. Tatan Y, Arvas A, Demir G, Alikai folu M, Gür E, Kiray E. Influence of Bacillus Calmette-Guèrin vaccination at birth and 2 months old age on the peripheral blood T-cell subpopulations [gamma/delta and alpha-beta T cell]. *Pediatr Allergy Immunol* 2005; 16: 624-9.
11. Dieli F, Troye-Blomberg M, Ivanyi J, Fournie JJ, Bonneville M, Peyrat MA, Sireci G, Salerno A. $V\gamma 9/V\delta 2$ T lymphocytes reduce the viability of intracellular Mycobacterium tuberculosis. *Eur J Immunol* 2000; 30: 1512-9.
12. Dieli F, Caccamo N, Meraviglia S, Ivanyi J, Sireci G, Bonanno CT, et al. Reciprocal stimulation of cd T cells and dendritic cells during the anti-mycobacterial immune response. *Eur J Immunol* 2004; 34: 3227-35.
13. Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS, et al. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum* 1988; 3: 315-24.
14. Antoni C, Braun J. Side effects of anti-TNF therapy: current knowledge. *Clin Exp Rheumatol* 2002; 20: S152-7.
15. Ehlers S. Role of tumour necrosis factor (TNF) in host defence against tuberculosis: implications for immunotherapies targeting TNF. *Ann Rheum Dis* 2003; 62: ii 37.
16. Stenger S. Immunological control of tuberculosis: role of tumour necrosis factor and more. *Annals of the Rheumatic Diseases* 2005; 64 (Supplement 4): iv 24-iv 28.
17. Salerno A, Dieli F. Role of gamma-delta T lymphocyte in immune response in humans and mice. *Crit Rev Immunol* 1998; 18: 327-57.
18. Burk MR, Mori L, De Libero G. Human $V\gamma 9-V\delta 2$ cells are stimulated in a cross-reactive fashion by a variety of phosphorylated metabolites. *Eur J Immunol* 1995; 25: 2052-8.
19. Olive C, Gatenby PA, Serjeantson SW. Persistence of gamma/delta T cell oligoclonality in the peripheral blood of rheumatoid arthritis patients. *Eur J Immunol* 1994; 72: 7-11.
20. Bank I, Cohen L, Moullem M, Farfel Z, Grossman E, Ben-Nun A. $\gamma\delta$ T cell subsets in patients with arthritis and chronic neutropenia. *Ann Rheum Dis* 2002; 61: 438-43.
21. Dieli F, Cacciamo N, Meraviglia S, Ivanyi J, Sireci G, Bonanno C, et al. Reciprocal stimulation of $\gamma\delta$ T cells and dendritic cells during the anti-mycobacterial immune response. *Eur J Immunol* 2004; 34: 3227-35.
22. Caccamo N, Sireci G, Meraviglia S, Dieli F, Ivanyi J, Salerno A. Gammadelta T cells condition dendritic cells in vivo for priming pulmonary CD8 T cell responses against Mycobacterium tuberculosis. *Eur J Immunol* 2006; 36: 2681-90.
23. Dieli F, Sireci G, Caccamo N, Di Sano C, Titone L, Romano A, et al. Selective depression of interferon-gamma and granulysin production with increase of proliferative response by $V\gamma 9/V\delta 2$ T cells in children with tuberculosis. *J Infect Dis* 2002; 186: 1835-9.